

# ***The Role and Clinical Application of Vaginal Microbiota in the Progression of Cervical Diseases***

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**Abstract.** Increasing evidence indicates that the vaginal microbiota (VMB) is closely associated with the development and progression of cervical diseases. Alterations in microbial composition have been observed across different stages of human papillomavirus (HPV) infection, cervical intraepithelial neoplasia (CIN), and cervical cancer. In healthy individuals, the VMB is typically dominated by *Lactobacillus* species, which contribute to the maintenance of an acidic environment and local immune balance. In contrast, vaginal dysbiosis is characterized by a reduction in *Lactobacillus* and an increased abundance of anaerobic bacteria, accompanied by elevated pH, disruption of epithelial barriers, and persistent inflammatory responses. Clinical and sequencing studies have reported that enrichment of specific bacterial genera, including *Sneathia* and *Fusobacterium*, is more frequently detected in high-grade cervical lesions, suggesting a potential association with disease severity. These microbial shifts may facilitate persistent HPV infection and promote lesion progression through multiple biological pathways. In addition, emerging evidence suggests that modulation of the VMB, particularly through probiotic supplementation with *Lactobacillus crispatus*, may support HPV clearance and improve microbial stability in early disease stages. By integrating findings from recent studies, this review outlines the dynamic changes of key microbial taxa during cervical disease progression and discusses their potential clinical relevance. These observations provide a quantitative reference for future investigations and highlight the possible value of microbiome-based strategies in cervical disease risk assessment and management.

**Keywords:** Vaginal microbiota, HPV infection, cervical intraepithelial neoplasia, cervical cancer

## **1. Introduction**

Cervical cancer is one of the most common malignant tumors among women worldwide. Persistent infection with high-risk human papillomavirus (HPV) has been established as a necessary etiological factor [1]. However, HPV infection alone is insufficient to cause cervical cancer, as the

host microenvironment plays a crucial role in its progression [2]. In recent years, the vaginal microbiota (VMB), as a key component of the cervical local environment, has emerged as a focal point in the study of the pathogenesis of cervical diseases [3].

Increasing evidence suggests that variations in microbial composition are associated with different outcomes of HPV infection and with the development of cervical lesions [3]. In healthy women, the VMB is typically characterized by the dominance of *Lactobacillus* species, which contribute to vaginal acidity and epithelial barrier integrity through the production of lactic acid and other antimicrobial metabolites. In contrast, disruption of this microbial balance is often accompanied by a reduction in *Lactobacillus* abundance and an expansion of anaerobic bacteria. Such alterations are associated with elevated vaginal pH, increased microbial diversity, and sustained inflammatory responses, conditions that may favor persistent HPV infection and lesion progression [3].

Accumulating studies have examined changes in the vaginal microbial community across the spectrum of cervical disease, ranging from cervicitis and cervical intraepithelial neoplasia (CIN) to invasive cervical cancer. These investigations have reported consistent shifts in the relative abundance of specific bacterial taxa, as well as potential mechanistic links between microbial dysbiosis and epithelial transformation. In this context, the present review synthesizes recent findings on VMB alterations during cervical disease progression and discusses their possible clinical implications, including risk stratification, microbiome-based biomarkers, and microbiota-targeted interventions.

## 2. The VMB as a key regulator of cervical disease

The VMB plays a central regulatory role in cervical disease. As a dynamic ecosystem integrating microbial communities, endocrine factors, anatomy, and mucosal immunity [4], its transition from health to dysbiosis critically shapes the outcome of HPV infection [5]. Understanding VMB dynamics has therefore become essential for elucidating cervical carcinogenesis, identifying biomarkers, and developing novel interventions.

As shown in Table 1, we summarize the main features of the vaginal microenvironment in both healthy and dysbiotic states. A healthy VMB is characterized by low microbial diversity and dominance of *Lactobacillus* species, especially *Lactobacillus crispatus*, *Lactobacillus gasseri*, and *Lactobacillus jensenii* [6]. These species maintain a low vaginal pH (3.8–4.5) through lactic acid production and secrete antimicrobial factors such as H<sub>2</sub>O<sub>2</sub> and bacteriocins. Among Community State Types (CSTs), CST I (*L. crispatus*) is considered the most stable and protective because of its efficient D-lactic acid production.

Table 1. Comparison of vaginal microenvironment between healthy and dysbacteriotic states

Feature	Health Status	Transition Status	Dysbiosis Status
Dominant bacterial genus/species	<i>L. crispatus</i> , <i>L. gasseri</i> , <i>L. jensenii</i>	<i>L. iners</i>	<i>Gardnerella</i> , <i>Prevotella</i> , <i>Famyhessea</i> , <i>Sneathia</i>
Common CSTs	CST I, II, V	CST III	CST IV

Table 1. (continued)

Typical pH value	<4.5	>4.4	>4.5
Microbial diversity	low	low	high
Key metabolites	D-Lactic acid, hydrogen peroxide, bacteriocin	L-Lactic acid	Amines, sialidase, short chain fatty acids
Mucosal barrier integrity	Complete, thick mucus layer	Poor mucus layer function	Damaged, mucous layer degraded
Immune characteristics	Steady state, local immune tolerance	Relatively high baseline inflammatory state	Pro-inflammatory, elevated cytokine levels (IL-1 $\beta$ , TNF - $\alpha$ )
Related clinical outcomes	Healthy, resistant to infection	Sub optimal/fragile state, prone to transition to CST IV, increased risk of persistent HPV infection and CIN	Bacterial vaginosis, aerobic vaginitis, increased risk of HPV infection, and increased risk of premature birth, miscarriage, and pelvic inflammatory disease

Dysbiosis is characterized by loss of *Lactobacillus* dominance and an increase in anaerobic species, forming CST IV. Studies have shown that this microbial configuration commonly includes genera such as *Gardnerella*, *Prevotella*, *Atopobium*, *Sneathia*, and *Fusobacterium* [7,8]. The presence of these taxa is associated with elevated vaginal pH and has been reported in conditions such as bacterial vaginosis (BV) and aerobic vaginitis (AV), as well as in women with increased susceptibility to HPV infection, adverse pregnancy outcomes, and pelvic inflammatory disease [9]. Although *L. iners* (CST III) is classified within the *Lactobacillus* genus, accumulating evidence suggests that this state represents an unstable transitional microbiota. Compared with other *Lactobacillus* species, *L. iners* exhibits limited protective capacity and lacks the ability to produce hydrogen peroxide and several key biosynthetic products [10,11]. VMB communities dominated by *L. iners* have therefore been frequently observed in association with vaginal dysbiosis, persistent HPV infection, and an increased risk of CIN [3,12]. These findings indicate that species-level resolution may be more informative than genus-level classification when evaluating cervical disease risk. Vaginal dysbiosis should be regarded as an active pathogenic process. CST IV taxa degrade the cervical mucus barrier, form biofilms, and generate inflammatory metabolites [13,14]. These mechanisms highlight the need for therapeutic strategies that not only replenish protective *Lactobacillus* species but also disrupt pathogenic biofilms [15].

The progression of cervical disease parallels a predictable gradient of microbial deterioration. Cervicitis, often idiopathic, is strongly associated with BV-like dysbiosis. BV-associated bacteria (*Gardnerella*, *Prevotella*, *Atopobium*) correlate with inflammation that may facilitate initial HPV infection. Meta-analyses show dysbiosis increases the risk of acquiring HPV by 33–43% and persistent infection by ~14% [16]. Women with BV have a 1.8–3.4-fold higher risk of persistent high-risk HPV, and *Lactobacillus*-depleted or *L. iners*-dominated VMB shows a 2–4-fold greater likelihood of HPV positivity [17,18]. Several anaerobes (*Gardnerella*, *Prevotella*, *Sneathia*, *Fusobacterium*) serve as potential biomarkers for HPV infection.

As cervical lesions progress from low-grade squamous intraepithelial lesion (LSIL/CIN1) to high-grade squamous intraepithelial lesions (HSIL/CIN2-3), VMB diversity increases while *Lactobacillus* abundance declines [19]. High-grade lesions are particularly associated with CST IV

and enrichment of *Sneathia*, *Prevotella*, and *Gardnerella*, which promote inflammation, HPV E6/E7 expression, and viral genome integration, accelerating malignant transformation [20,19]. Even after surgical treatment, dysbiosis often persists, explaining high recurrence rates. In cervical cancer, dysbiosis peaks: alpha diversity markedly increases, *Lactobacillus* species are depleted, and pathogenic anaerobes such as *Fusobacterium*, *Sneathia*, *Porphyromonas*, and *Peptostreptococcus* become dominant [21,22]. Certain taxa, including *Sneathia sanguinegens* and *Peptostreptococcus anaerobius*, can effectively distinguish HSIL from LSIL [3].

These changes have been repeatedly validated across multiple independent studies. To visually demonstrate this quantitative process, Table 2 integrates data from several investigations, illustrating the trends in the average relative abundance of key microbial taxa across different stages of cervical disease. Although these reference values are not absolute due to variations in study design, populations, and analytical methods, they consistently point toward a similar biological trajectory and highlight the substantial potential of microbiome-based diagnostics. This clear and directional pattern of microbial community succession from health to malignancy strongly suggests a co-evolutionary relationship between microbial communities and host tissues.

Table 2. Key vaginal microenvironment characteristics at different stages of cervical disease development

Microbe	Health (HPV-)	Asymptomatic HR HPV+	LSIL (CIN1)	HSIL (CIN2/3)	Cervical cancer
<i>L. crispatus</i>	50-80	30-60	10-30	< 5	< 1
<i>L. iners</i>	10-30	20-40	20-50	10-30	5-20
<i>Gardnerella vaginalis</i>	< 1	1-10	5-20	10-30	10-25
<i>Prevotella</i>	< 0.5	< 2	2-10	5-15	5-20
<i>Sneathia</i>	< 0.1	< 1	1-5	5-20	8-25
<i>Fusobacterium</i>	< 0.01	< 0.1	< 0.5	1-5	2-10
<i>Atopobium vaginae</i>	< 0.5	< 2	2-8	5-15	5-15
$\alpha$ -diversity (Shannon index)	Low (1.0-1.5)	Low (1.2-1.8)	Middle (1.8-2.5)	High (2.5-3.5)	High (2.8-4.0)
Key metabolite characteristics	Lactic acid dominance	Start transitioning to short chain fatty acids (SCFAs)	SCFAs, Increased biogenic amines	High level SCFAs, altered lipid/amino acid profile	Complex metabolic dysbiosis, significant changes in biogenic amines, lipids, etc

### 3. Vaginal virome and the microbial continuum

In addition to bacteria, the VMB also harbors a complex community of viruses, collectively referred to as the virome, which is predominantly composed of bacteriophages (phages, viruses that infect bacteria) [23]. Phages serve as key regulators of bacterial communities. Phages specific to *Lactobacillus* species are associated with *Lactobacillus*-dominated microbiota and may influence community stability [24]. Moreover, under dysbiotic conditions, certain phages show positive correlations with BV-associated taxa such as *Gardnerella*, *Prevotella*, and *Atopobium*, and these

cross-domain interactions have been linked to genital tract inflammation [25,26]. This suggests that phages may play a critical role in shaping pathogenic bacterial consortia that drive disease processes. Other viruses, including anelloviruses, are also present in the vagina. Their abundance correlates with genital tract inflammation, further adding complexity to the role of the virome in cervical health [27,28].

Meanwhile, cervical health should not be viewed in isolation, as it forms an integral part of an interconnected reproductive tract ecosystem. Earlier concepts regarded the upper reproductive tract including the uterus and fallopian tubes as a sterile environment [29]. With the application of high-throughput sequencing techniques, microbial DNA has now been detected in the endometrium in the proximal region of the reproductive tract. Analyses of these microbial communities indicate that their composition frequently shares characteristics with the cervical and VMB [30]. These observations support the concept of a microbial continuum along the female reproductive tract, in which microorganisms from the lower genital tract may ascend and establish a dynamic ecological gradient. In this framework, alterations of the VMB are not restricted to local cervical effects but may influence the microbial balance of the endometrium. Several studies have reported associations between lower genital tract dysbiosis and endometrial conditions, including chronic endometritis and infertility, and have further suggested possible links to gynecologic malignancies such as endometrial and ovarian cancers [31]. Consistent with this view, inflammatory signatures and bacterial taxa implicated in cervical lesion progression, particularly *Gardnerella* and *Prevotella*, have also been identified in endometrial samples from women diagnosed with infertility or endometritis [32]. Rather than remaining confined to the cervix, these microorganisms and the inflammatory responses they promote may extend across anatomical boundaries, thereby influencing disease susceptibility throughout the reproductive tract [33].

The above evidence suggests that interventions aimed at restoring vaginal health may confer profound benefits for overall reproductive health. Conversely, persistent vaginal dysbiosis poses a systemic risk to the entire female reproductive tract.

#### 4. Clinical frontiers: diagnosis, treatment and prevention

##### 4.1. VMB as a predictive biomarker

The VMB is emerging as a valuable clinical tool for improving diagnosis, risk prediction, and treatment of HPV-related cervical disease. Although HPV testing and cytology provide high sensitivity, their limited specificity leads to unnecessary colposcopies and overtreatment [34]. VMB-based biomarkers offer a promising strategy for refined risk stratification. Specific microbial signatures, such as increased *Stenotrophomonas*, *Streptococcus*, and *Pseudomonas*, along with reduced *Faecalibacterium* and *Bifidobacterium*, have been proposed as indicators of HSIL [35]. Machine-learning models, including a random forest classifier built on 33 bacterial taxa, highlight *L. iners* as a major predictor of CIN severity [36]. Additional species combinations, such as *Prevotella bivia* and *Porphyromonas uenonis*, show potential in predicting persistent HPV infection or CIN2+ [37]. Ultimately, VMB profiling may enable accurate triage of HPV-positive women by identifying those at genuine risk of progression who require immediate evaluation.

##### 4.2. Therapeutic modulation of the microbiome: probiotics, prebiotics, and synbiotics

Microbiome-modulating therapies represent another frontier for clinical intervention. Several probiotic studies suggest potential benefits for HPV clearance and CIN regression. Daily intake of

Lactobacillus casei doubled LSIL regression rates compared with controls [38]. Oral L. rhamnosus and L. reuteri reduced abnormal cytology, though without a clear effect on HR-HPV clearance [39]. Intravaginal transplantation of L. crispatus chen-01 showed promising results, including reduced HPV viral load and restoration of healthy VMB composition [40]. A large Bayesian network meta-analysis evaluating eight therapeutic modalities found that combining interferon with Lactobacillus vaginal capsules or with Fuanning produced the strongest HR-HPV clearance effects, outperforming interferon alone [41].

Prebiotics, such as dietary fiber and inulin, have been reported to facilitate the growth of beneficial vaginal bacteria and may contribute to reduced HPV risk. Synbiotics, which integrate probiotics and prebiotics, are likewise under active investigation. Recent consensus recommendations emphasize strain-specific approaches, particularly the use of L. crispatus and L. rhamnosus, tailored to clinical goals such as HPV clearance and CIN regression [42].

## 5. Conclusion

Accumulating evidence indicates that alterations in the VMB are closely associated with the development and progression of cervical disease. Rather than acting as a static background, the vaginal microbial ecosystem undergoes measurable shifts during HPV infection, cervical intraepithelial neoplasia, and invasive cervical cancer. In health, Lactobacillus-dominated communities help maintain an acidic environment and support local immune equilibrium. Disruption of this microbial structure is commonly accompanied by expansion of anaerobic bacteria, increased vaginal pH, and enhanced inflammatory activity, conditions that have been linked to persistent HPV infection and epithelial transformation. Specific bacterial taxa, including *Sneathia* and *Fusobacterium*, have been repeatedly observed at higher abundance in high-grade cervical lesions, suggesting a potential role as indicators of disease progression. These microbial signatures may complement existing screening strategies by providing additional biological context for risk assessment. Beyond diagnosis, insights into vaginal microbiota dynamics also open opportunities for intervention. Approaches aimed at restoring microbial balance, particularly those involving defined Lactobacillus strains such as L. crispatus, represent a promising area of ongoing investigation. Continued longitudinal studies and well-designed clinical trials will be essential to clarify their effectiveness and to determine how microbiome-based strategies can be integrated into cervical disease prevention and management.

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## Conflict of interest

There is no conflict of interest.

## References

- [1] Perkins R.B., Wentzensen N., Guido R.S., Schiffman M. (2023) Cervical cancer screening: A review. *JAMA*, 330, 547-558.
- [2] Yuan Y., Cai X., Shen F., Ma F. (2021) Hpv post-infection microenvironment and cervical cancer. *Cancer Lett*, 497, 243-254.
- [3] Mitra A., MacIntyre D.A., Marchesi J.R., et al. (2016) The vaginal microbiota, human papillomavirus infection and cervical intraepithelial neoplasia: What do we know and where are we going next? *Microbiome*, 4, 58.
- [4] Cui M., Wu Y., Liu Z., Liu Y., Fan L. (2025) Advances in the interrelated nature of vaginal microecology, hpv infection, and cervical lesions. *Front Cell Infect Microbiol*, 15, 1608195.
- [5] Schellekens H.C.J., Schmidt L.M.S., Morre S.A., van Esch E.M.G., de Vos van Steenwijk P.J. (2025) Vaginal microbiota and local immunity in hpv-induced high-grade cervical dysplasia: A narrative review. *Int J Mol Sci*, 26,
- [6] Valeriano V.D., Lahtinen E., Hwang I.-C., et al. (2024) Vaginal dysbiosis and the potential of vaginal microbiome-directed therapeutics. *Frontiers in Microbiomes*, 3,
- [7] Witkin S.S., Mendes-Soares H., Linhares I.M., et al. (2013) Influence of vaginal bacteria and d- and l-lactic acid isomers on vaginal extracellular matrix metalloproteinase inducer: Implications for protection against upper genital tract infections. *mBio*, 4,
- [8] Ma Z.S., Li L. (2017) Quantifying the human vaginal community state types (csts) with the species specificity index. *PeerJ*, 5, e3366.
- [9] Chen X., Lu Y., Chen T., Li R. (2021) The female vaginal microbiome in health and bacterial vaginosis. *Front Cell Infect Microbiol*, 11, 631972.
- [10] Macklaim J.M., Gloor G.B., Anukam K.C., Cribby S., Reid G. (2011) At the crossroads of vaginal health and disease, the genome sequence of *lactobacillus iners* ab-1. *Proc Natl Acad Sci U S A*, 108 Suppl 1, 4688-4695.
- [11] Zheng N., Guo R., Wang J., Zhou W., Ling Z. (2021) Contribution of *lactobacillus iners* to vaginal health and diseases: A systematic review. *Front Cell Infect Microbiol*, 11, 792787.
- [12] Usyk M., Zolnik C.P., Castle P.E., et al. (2020) Cervicovaginal microbiome and natural history of hpv in a longitudinal study. *PLoS Pathog*, 16, e1008376.
- [13] Castro J., Machado D., Cerca N. (2019) Unveiling the role of *gardnerella vaginalis* in polymicrobial bacterial vaginosis biofilms: The impact of other vaginal pathogens living as neighbors. *ISME J*, 13, 1306-1317.
- [14] Morrill S., Gilbert N.M., Lewis A.L. (2020) *Gardnerella vaginalis* as a cause of bacterial vaginosis: Appraisal of the evidence from in vivo models. *Front Cell Infect Microbiol*, 10, 168.
- [15] Ma X., Wang X., Ye S., et al. (2022) Biofilm and pathogenic factor analysis of *gardnerella vaginalis* associated with bacterial vaginosis in northeast china. *Front Microbiol*, 13, 1033040.
- [16] Brusselaers N., Shrestha S., van de Wijgert J., Verstraeten H. (2019) Vaginal dysbiosis and the risk of human papillomavirus and cervical cancer: Systematic review and meta-analysis. *Am J Obstet Gynecol*, 221, 9-18 e18.
- [17] Kazlauskaitė J., Zukiene G., Rudaitis V., Bartkevičiūtė D. (2025) The vaginal microbiota, human papillomavirus, and cervical dysplasia-a review. *Medicina (Kaunas)*, 61,
- [18] Liang Y., Chen M., Qin L., Wan B., Wang H. (2019) A meta-analysis of the relationship between vaginal microecology, human papillomavirus infection and cervical intraepithelial neoplasia. *Infect Agent Cancer*, 14, 29.
- [19] Peng Y., Tang Q., Wu S., Zhao C. (2025) Associations of atopobium, garderella, megasphaera, prevotella, sneathia, and streptococcus with human papillomavirus infection, cervical intraepithelial neoplasia, and cancer: A systematic review and meta-analysis. *BMC Infect Dis*, 25, 708.
- [20] Curty G., de Carvalho P.S., Soares M.A. (2019) The role of the cervicovaginal microbiome on the genesis and as a biomarker of premalignant cervical intraepithelial neoplasia and invasive cervical cancer. *Int J Mol Sci*, 21,
- [21] Li X., Wu J., Wu Y., et al. (2023) Imbalance of vaginal microbiota and immunity: Two main accomplices of cervical cancer in chinese women. *Int J Womens Health*, 15, 987-1002.
- [22] Hurst R., Brewer D.S., Gihawi A., Wain J., Cooper C.S. (2024) Cancer invasion and anaerobic bacteria: New insights into mechanisms. *J Med Microbiol*, 73,
- [23] Huang L., Guo R., Li S., et al. (2024) A multi-kingdom collection of 33, 804 reference genomes for the human vaginal microbiome. *Nat Microbiol*, 9, 2185-2200.
- [24] Hugerth L.W., Krog M.C., Vomstein K., et al. (2024) Defining vaginal community dynamics: Daily microbiome transitions, the role of menstruation, bacteriophages, and bacterial genes. *Microbiome*, 12, 153.
- [25] Deng Z.L., Gottschick C., Bhuju S., et al. (2018) Metatranscriptome analysis of the vaginal microbiota reveals potential mechanisms for protection against metronidazole in bacterial vaginosis. *mSphere*, 3,
- [26] Ali A., Jorgensen J.S., Lamont R.F. (2022) The contribution of bacteriophages to the aetiology and treatment of the bacterial vaginosis syndrome. *Fac Rev*, 11, 8.

- [27] Kaelin E.A., Skidmore P.T., Laniewski P., et al. (2022) Cervicovaginal DNA virome alterations are associated with genital inflammation and microbiota composition. *mSystems*, 7, e0006422.
- [28] Li Y., Cao L., Han X., et al. (2023) Altered vaginal eukaryotic virome is associated with different cervical disease status. *Virol Sin*, 38, 184-197.
- [29] Gholiof M., Adamson-De Luca E., Wessels J.M. (2022) The female reproductive tract microbiotas, inflammation, and gynecological conditions. *Front Reprod Health*, 4, 963752.
- [30] Canha-Gouveia A., Perez-Prieto I., Rodriguez C.M., et al. (2023) The female upper reproductive tract harbors endogenous microbial profiles. *Front Endocrinol (Lausanne)*, 14, 1096050.
- [31] Toson B., Simon C., Moreno I. (2022) The endometrial microbiome and its impact on human conception. *Int J Mol Sci*, 23,
- [32] Liang J., Li M., Zhang L., et al. (2023) Analysis of the microbiota composition in the genital tract of infertile patients with chronic endometritis or endometrial polyps. *Front Cell Infect Microbiol*, 13, 1125640.
- [33] Govorov I., Komlichenko E., Ulrich E., et al. (2025) The microbiome in endometrial cancer: Vaginal milieu matters. *Front Med (Lausanne)*, 12, 1533344.
- [34] Force U.S.P.S.T., Curry S.J., Krist A.H., et al. (2018) Screening for cervical cancer: Us preventive services task force recommendation statement. *JAMA*, 320, 674-686.
- [35] Chao X., Wang L., Wang S., et al. (2021) Research of the potential vaginal microbiome biomarkers for high-grade squamous intraepithelial lesion. *Front Med (Lausanne)*, 8, 565001.
- [36] Lee Y.H., Kang G.U., Jeon S.Y., et al. (2020) Vaginal microbiome-based bacterial signatures for predicting the severity of cervical intraepithelial neoplasia. *Diagnostics (Basel)*, 10,
- [37] Chao X., Sun T., Wang S., et al. (2020) Research of the potential biomarkers in vaginal microbiome for persistent high-risk human papillomavirus infection. *Ann Transl Med*, 8, 100.
- [38] Verhoeven V., Renard N., Makar A., et al. (2013) Probiotics enhance the clearance of human papillomavirus-related cervical lesions: A prospective controlled pilot study. *Eur J Cancer Prev*, 22, 46-51.
- [39] Ou Y.C., Fu H.C., Tseng C.W., et al. (2019) The influence of probiotics on genital high-risk human papilloma virus clearance and quality of cervical smear: A randomized placebo-controlled trial. *BMC Womens Health*, 19, 103.
- [40] Liu Y., Zhao X., Wu F., et al. (2024) Effectiveness of vaginal probiotics *lactobacillus crispatus chen-01* in women with high-risk hpv infection: A prospective controlled pilot study. *Aging (Albany NY)*, 16, 11446-11459.
- [41] Tang Y., Tong Q., Dai N., Xu C. (2023) Evaluation of the clinical efficacy of vaginal treatment options for persistent high-risk human papillomavirus infection after excisional treatment of cervical high-grade squamous intraepithelial lesions: A systematic review and bayesian network meta-analysis. *Virol J*, 20, 47.
- [42] Gutierrez Salmean G., Delgadillo Gonzalez M., Rueda Escalona A.A., Leyva Islas J.A., Castro-Eguiluz D. (2024) Effects of prebiotics, probiotics, and synbiotics on the prevention and treatment of cervical cancer: Mexican consensus and recommendations. *Front Oncol*, 14, 1383258.